

# Cytogenetics of *Sarcococca orientalis*

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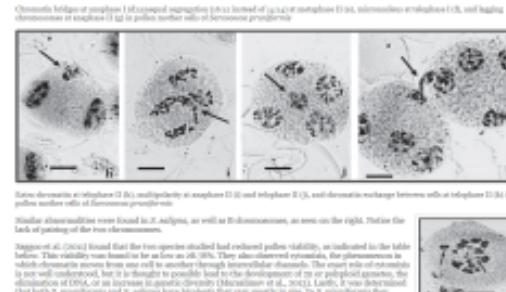
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## Introduction and Literature Review

*Sarcococca*, or boxwood, is a genus of 16 species of evergreen usually shrubby native primarily to Asia (Chen, 2001). As members of the Buxaceae, they are close relatives to common temperate plants including hollies (Ilex), boxwoods (Buxus), and magnolias (Magnolia). All species are dioecious, though some are monoecious, though they are reported for their largely winter flowers and shade adaptability (The Plant List, 2010). In my personal experience, few species (S. confusa, S. angustifolia, S. hookeriana, and S. mollis) are seen in America.

### Cytogenetics

*Sarcococca* has been well-explored cytologically. Of the six species, only four have reported chromosome numbers (S. confusa, S. angustifolia, S. mollis, and S. orientalis), of which all are unreported taxa and may be present in the literature. The first report of a base chromosome number of  $n = 14$  was by S. S. Sankaran in 1971, followed by a report of  $n = 14$  in 1977 by Chen et al. (1977). A more recent report of  $n = 14$  was by the identification of chromosome numbers in five species, the only other cytological work in 2011 paper in which the authors observed counts in S. angustifolia and related it to pollen viability (Rengaraju et al., 2011). In this page, the authors found several instances of mitotic chromosomes, some of which are presented below.



Chromosome bridges or anaphase I of unequal separation (d,e,f,g) in pollen mother cells of *Sarcococca* D-14, micrographs: a=anaphase I, b=metaphase II, c=telophase II, d=anaphase II, e=gene conversion in pollen mother cells of *Sarcococca* D-14.



Later division in metaphase II (h), anaphase II (i), and telophase II (j), and de-nucleus exchange between cells in telophase II (k) in pollen mother cells of *Sarcococca* D-14.

Binucleate pollen mother cells were found in S. angustifolia, an cell at 10 chromosomes, as seen on the right. Photo is the last of pairing of the two chromosomes.

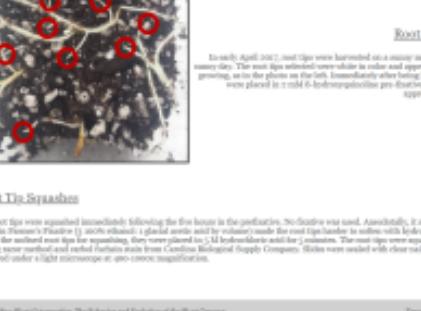
Rengaraju et al. (2011) found that the four species studied had reduced pollen viability, as indicated in the table below. This variability was found to be as low as 20%. They also observed cytology, the phenomenon in which chromosomes move from cell to another through nuclear通道. The exact role of re-nucleation is not completely understood, but it is believed to be involved in the development of pollen grains. It was determined that S. confusa, S. angustifolia, and S. mollis have binucleate cells that vary greatly in size. In S. confusa, it was found that 3% of the cells had 4 and 4-fold binucleate as compared to 1, while they found three exceptionally small binucleate of 14 total (in S. mollis).

	Cytology			Pollen grains		
	PMC's %	No. of PMC's involved	Mitotic stages/ages	Average Binucleate (%)	Size (μm)	BF
<i>Sarcococca angustifolia</i> Lindl.	—	—	—	28.78	14.46±11.85	0.5
Kengs, Sikkim	—	—	—	28.46±11.25	17.5	0.5
Kengs, eastern Tibet	1.80	2	T-II	87.51	11.46±11.35	0.6
<i>S. mollis</i> (D. Don) Muell.-Arg. Pouch. Naldehra	—	—	—	71.83	28.36±20.66	0.5
				20.45±14.00	16.17	

Pollen viability and cytology in *Sarcococca angustifolia* and *S. mollis*. PMC - pollen mother cell, BF - relative frequency of small and large pollen grains.

### Apoliosis

Considering the amount of mitotic abnormalities and the low pollen viability found in both *Sarcococca* species, it may not be surprising that at least one species in the genus has an apopotic pathway to a certain extent. Apoptosis is a programmed cell death, involving the expression of pro-apoptotic genes and the execution of caspases. In plants, apoptosis is a method of escape from other cell damage or damage in polyplids (Carmi, 1997) and it is often used for removing damaged plant cells and increasing efficiency of pollination in self-pollinators. However, the only reported example in the literature does not have a known pathway, and the only known pathways are digital.



Two seedlings from a single seed in both *Sarcococca angustifolia* and *S. mollis* hybrids.

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## Objective

As stated in the introduction, quantitative approach for a common chromosome in *Sarcococca*. However, because *Sarcococca* is more common in higher plants and the only known plant in *Sarcococca* is diploid, I wanted to investigate chromosome number in a species with tetraploid chromosome number. This could provide insight into the evolutionary history of the genus. For example, it may be possible to determine the ploidy level of the species by observing the number of chromosomes in the genome. If the genome is diploid, then the number of chromosomes will be even. If the genome is tetraploid, then the number of chromosomes will be odd. This may help explain the difference in the number of chromosomes in polyploids and diploids.

Moreover, the only reported example in the literature does not have a known pathway, and the only known pathways are digital.

*Sarcococca orientalis* is a woody shrublet candidate for this study. It has an unknown chromosome number and large, dark leaves that may be Sarcococca species. While not a true indicator, larger leaves is a common feature in polyploids [Guzman and Lescano, 1990].

### Materials and Methods

#### Species Propagation

On January 19, 2011, cuttings approximately 6 inches in length were taken and dipped in Rootone (2,4-dichlorophenoxyacetic acid, 1000 ppm) for 30 seconds and 1 before being stuck. The cutting was placed under mist until rooting in an automatic misting unit (not shown) and then placed in a pot with hydroponic soil.



Root Tip Harvest

In early April 2012, root tips were harvested on a rainy morning following a sunny day. The root tips were harvested in order and separated to be analyzed. As the photos on the left, immediately after being harvested, the tips were placed in 7 ml 0.06% hydroxyapatite pre-filtrate at 4 degrees C for approximately 3 hours.

#### Root Tip Squash

The root tips were squashed immediately following the first hour in the pre-filtrate. No fixative was used. Anecdotally, it seemed like 25 hours in Fixer's Fixative (1:1000 dilution) 1:1000 dilution 1:1000 made the root tips harder to subdue with hydrochloric acid. To reduce the number of tips for squashing, they were placed in 5.12 hydroxyapatite and 24 minutes. The root tips were squashed using the sliding zone method and crushed further with a Cytospin Centrifuge (Shandon Scientific). Roots were washed with clear cold water and observed under a light microscope in *in-situ* mode, magnification.

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## Stages of Mitosis

### Prophase

Above left: Various stages of chromosome condensation during prophase. Above right: The cell on the left is clearly in prophase, while others are in various stages from interphase (entire cell) to early prophase.

### Metaphase



Above: Chromosomes lined up at metaphase. A few chromosomes are slightly lagging. The photos on the left and right show chromosomes more densely aligned at the metaphase plate than in the center photo.

### Anaphase



Above: Chromatids being pulled apart during anaphase. In the photo on the left, some chromatids are still in the process of being separated, while in the photo on the right, all chromatids have clearly been separated.

### Telophase



Above left: The cell on the left has undergone anaphase, chromosomes have mostly decondensed, and a cell plate has formed. The cell on the right has undergone telophase, and the chromosomes have decondensed. A cell plate has yet to form, however. Also notice the metaphase at the top of the photo.

Above right: The binucleate cell has undergone anaphase and chromosomes have mostly decondensed. A cell plate has yet to form.

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This website contains my final project for PHGC101: Plant Cytogenetics at the University of Georgia. The website is a place of discussion for anyone interested in plant cytogenetics, specifically in *Sarcococca* species having reported chromosome numbers. Of particular interest is the presence of specific species in the genera. I explore various and likely to related species using more advanced ploidy levels.

Please feel free to contact me if you have any questions or comments.

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## Chromosome Number



Above: Two pictures of a cell with varying condensations in heterochromosomes.  $2n = 28$  in *S. orientalis*.



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