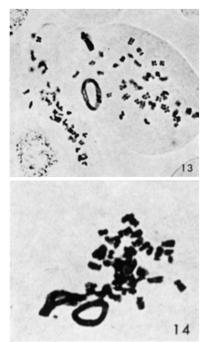
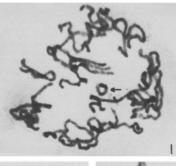
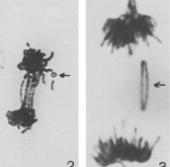
# **Ring chromosomes**

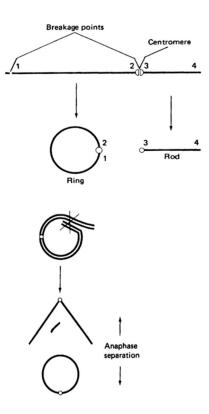
Review by Yu, 2018

- Arise through double breakage, one near the end, the other at the centromere
- Can also be formed by a crossover within a duplication



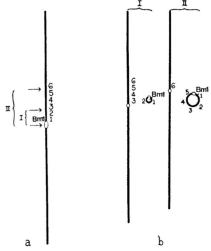






Ring chromosomes in tobacco Gerstel & Burns, 1967

Ring chromosomes in onion, Gohil & Kaul, 1983. Experentia 39: 1152-3.



# McClintock, 1938

Devised a system of deficient rods with complementary rings on chromosome 5 of maize

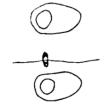
- Loss of a ring would give plants variegated for normal and brown midrib (whenever rings get lost)
- In a plant of genotype *bm1 bm1*, loss of a ring with *Bm1* results in brown tissue
- In a plant hemizygous for *bm1* (i.e., one homologue is deficient for the *bm1* allele), loss of a ring with *Bm1* results in dead tissue
- In a homozygous state, the deficiency is lethal.
- The rings were not transmitted through the gametophyte

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Small rings behave differently than large rings:

- Lost more frequently (right: lag behind, and don't get included in a nucleus)
- Change in size less frequently
- Reduced frequency of double-sized rings
- Increase in number 2 rings go to the same nucleus

Ring size	Freq. double-sized rings
Same as rod	15-20%
$^{1}/_{10}$ as big	1%
$^{1}/_{25}$ as big	0.2%



#### McClintock, 1941

Ring chromosomes become unstable if sister-strand crossing over occurs

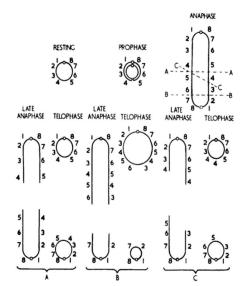
- increase in size through duplications
- decrease in size through deletions
- can be lost
- can increase in number
- A = breakage into equal halves followed by fusion
- B = breakage into unequal parts followed by fusion (altered size)
- C = breakage into unequal halves followed by fusion (similar size, but altered genetic content)

# **Breakage-Fusion-Bridge cycles**

#### McClintock, 1941; review by Jones, 2005

2 main types:

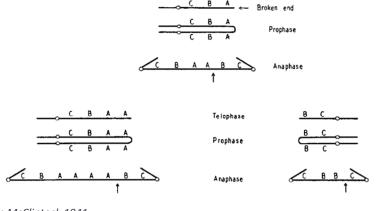
- chromosome (breakage of ring chromosomes in somatic tissues)
- chromatid



#### Chromosome breakage fusion bridge cycles:

When a chromosome with its end broken off replicates, its ends may fuse.

- This leads to the formation of a bridge during anaphase.
- The bridge breaks, and the cycle gets repeated
- When the break takes place, the resulting chromosomes will either have duplicate or deficient material.



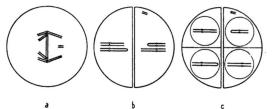
from McClintock 1941

# BFB in meiosis

# McClintock, 1938

To study BFB cycles in meiosis, need 2 broken ends in a cell

- Created these by a 4-strand double crossover within a large paracentric inversion in chromosome 4 of maize
- Gives a bridge and fragment after the first division, and 2 bridges during the second division



Breakage of the bridge takes place during anaphase or telophase II.

- About 25% of the microspores receive the broken chromosome.
- About 7.5% receive both the broken chromosome and the resulting fragment, thus resulting in a full genomic complement

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- Bridges form during the mitotic division of microgametogenesis

   This is chromatid breakage fusion bridge cycle
- At right is the first mitotic division of the microspore.
  - Note the bridge remaining between the daughter nuclei.
  - One of the fragment halves remains in the spindle, the other was included in the larger nucleus.

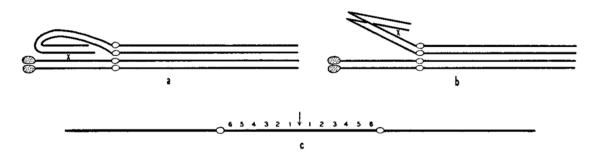
### **BFB** in the endosperm

#### McClintock, 1941

The difficulty in transmitting this to the next generation is that the inversion causes duplicatedeficient gametes.

- Consequently, McClintock devised a system that, even if a break occurred, the chromosome would not be deficient
- Used a chromosome 9 (with dominant genetic markers) that had an inverse duplication of in the short arm:

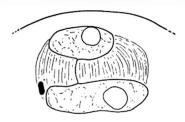
The duplicated fragment can either cross over with its homologue (a) or with itself (b), producing a dicentric chromosome with an inverted duplication in the middle (c):



(a) leads to the formation of a bridge and fragment at anaphase I

(b) leads to a bridge at anaphase II

• A break can still occur as indicated by the arrow, and still have intact chromosomes

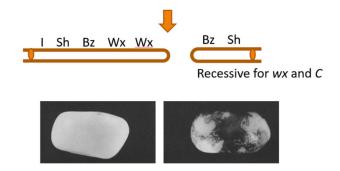


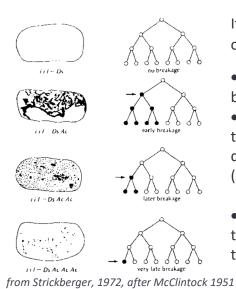
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#### McClintock, 1951

The markers make this easy to follow:

- The BFB cycle continued in the endosperm, but not in the embryo
- Leads to formation of variegated endosperm
  - Get all types of variegation, depending on when and where breaks occur:





If the cell is heterozygous for appropriate markers, the loss of genetic material results in a readily identifiable cell lineage

- This phenomenon does not occur in the embryos, as the broken ends heal rather than fuse.
- The original breakage is due to excision of the *Ds* transposable element. Increasing copy number of *Ac* decreases excision and breakage, leading to smaller spots (right) [*did not know @ time*].

• The maize kernel below was heterozygous for the *I* gene, the dominant allele of which inhibits color formation. Loss of the dominant allele leads to colored sectors (from

McClintock 1951)



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# **Rings and herbicide resistance**

# Palmer amaranth, the king of weeds, cripples new herbicides

Scientists in the US sound the alarm about a crop-smothering weed that is growing resistant to multiple herbicides

*by Melody M. Bomgardner* August 3, 2019 | A version of this story appeared in **Volume 97, Issue 31** 

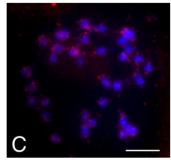
https://cen.acs.org/business/specialty-chemicals/Palmer-amaranth-king-weeds-cripples/97/i31



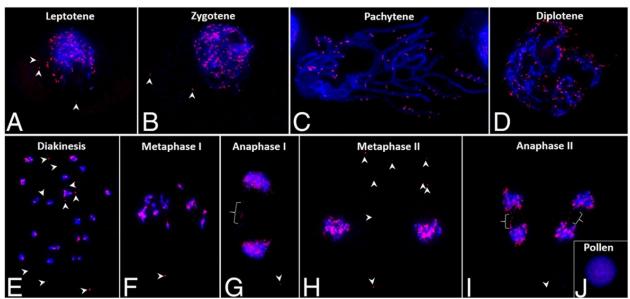
https://cropwatch.unl.edu/2017 /status-herbicide-resistantweeds-nebraska/

#### Koo et al, 2018a; Molin et al, 2020

- Resistant plants have 40-100 copies of EPSPS gene dispersed on all the chromosomes
- Increase genome size by 11%
- ~400-kb extrachromosomal circular DNA (eccDNA) that harbors the EPSPS gene and 58 other genes
- Transmitted by chromosome tethering during mitosis and meiosis



Gaines et al, 2009



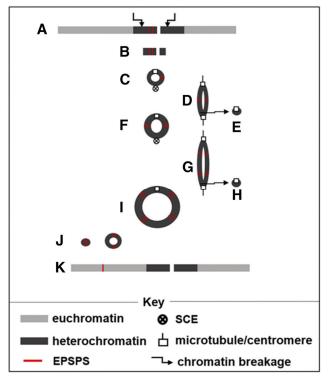
Tethered and unthethered eccDNAs during meiosis. From Koo et al, 2018

# Rings & BFB in herbicide tolerance Koo et al., 2018b

Water hemp, *Amaranthus tuberculatus*, is one of the weeds that has become resistant to glyphosate

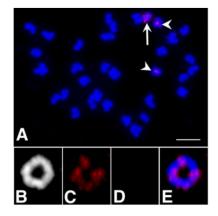
Photo shows a glyphosate resistant *A. tuberculatus* plant showed chromosome constitution of 2n = 32+1 circle (arrow)

FISH for EPSPS shows presence on two somatic chromosomes (arrow heads) and a cluster on the ring chromosome.





https://www.youtube.com/watch?v=Dg paxUkBeZA



#### Proposed model:

• Amplification of EPSPS-containing pericentromeric heterochromatin onto another chromosome

- Breakage leading to ring chromosome
- SSCOs lead to dicentric rings
- These break into different sized rings in continuing rounds of BFB cycles
- Some rings can reinsert into genome, forming new EPSPS loci