Inversions

Muller, 1916, working with Drosophila, found C (for crossover reducers) factors reduced or suppressed recombination in certain areas of the chromosome when they were in a heterozygous state

- By 1926, Sturtevant had compared the order of linked genes, and found the order had been inverted
- Inversions can be pericentric (around the centromere) or paracentric (beside the centromere)

A paracentric inversion



From US National Library of Medicine

X 👍 X

A pericentric inversion



Paracentric inversions



Crossover suppression

McClintock, 1931; 1933

Provided the cytological explanation as to why crossover was suppressed.

Nomenclature: Eg, the In 2a, S.6-L.6 inversion in maize

- In 2a In = inversion
 - 2 = chromosome 2
 - a = the first inversion found on this chromosome

S.6-L.6

- S.6 = breakpoint is at .6 of the distance from the centromere to the end of the short arm
- L.6 = breakpoint is at .6 of the distance from the centromere to the end of the long arm
- Working with a paracentric inversion on the short arm of chromosome 8:

Crossover inside loop (right):

- CO outside loop is inconsequential
- Formation of dicentric bridge and fragment at anaphase I The fragment is lost.



Only functional gametes are those from non-crossover strands. The rest are non-functional.

• This applies to odd numbers of crossovers

Double Crossovers in the Inversion Loops graphics from **Strickberger**, **1976**

1) 2-strand double crossover





All gametes are functional. Inversion cannot be detected unless have the appropriate markers.

2) 3-strand double crossover



Bridge and fragment seen at Ana I

• 50% sterility

• Of the surviving gametes, 50% will be recombinant

3) 4-strand double crossover



- 2 bridges & 2 fragments at Ana I
- All gametes formed are unviable

If one of the double crossovers occurs outside of the loop

Modified from McClintock, 1938

A. Get same configuration for a 1 or 3 strand double crossover

 From **Burnham, 1962**, after **McClintock, 1938** Ana I from Stevens & Bougourd. 1991. Heredity 66: 391-401 Ana II from Wang & Zhang. 2007. Plant Sci 172: 380-392

 Not affected by CO outside of inversion area



Single Crossover @ 1 or 2 \rightarrow bridge & fragment at anaphase I



Balanced: involves equal chromosome lengths on either side of the centromere

Unbalanced: involves unequal chromosome lengths on both sides of the centromere

- No bridges or fragments are formed •
- Half of the gametes (the recombinant ones) are duplicatedeficient





D C B

McClintock, 1931

Multiple inversions

Case 1: Nested inversions





The normal chromosome forms a small loop within the large loop.

Case 2: Overlapping inversions



Can be used to trace the history of inversions and reconstruct evolutionary trees. In this example, the possibilities are:

A to B to C or A \leftarrow B \rightarrow C or C \rightarrow B \rightarrow A

Paracentric inversions can be common in some organisms in natural populations

- By preventing recombination, isolate large blocks of genes of adaptive value
- Pericentric inversions source of heteromorphic arms in orthopteran insects

Role of inversions

Dobzhanski, with Drosophila:

Certain inversions or inversion combinations are adapted to particular ecological combinations. These differ with altitude and with seasons

Conclusions

- 1. Inversion complex has selective value
- 2. Population is in a state of flux
- 3. Heterozygotes have a greater selection value than either homozygote
- 4. Optimal size to inversions

Roles of inversions

- 1) Genes within inversions may diverge
- 2) ¿Could serve as initial barriers for speciation?

Structural variants - inversions

Huang & van der Knaap, 2011



Origin of inversions

Casals & Navarro, 2007

Model 1: Duplications lead to inversions



Model 2: Staggered breaks



Left: Ectopic recombination model

- Most accepted
- Duplications lead to inversions

Right: Staggered breaks model – eg., as might happen with transposon excision or radiation

• Inversion leads to duplication

Models are not mutually exclusive!

Reversing inversions

Schmidt et al, 2020

Because inversions act as recombination suppressors, genes in inverted areas are not amenable to breeding.

- In arabidopsis, chromosome 4 in Columbia has a paracentric inversion relative to Landsberg Erecta
- Using CRISPR to make simultaneous cuts, they were able to reverse the inversion with a 0.5% frequency.



Schwartz et al, 2020

Some corn inbreds have a 26.7 Mb or 75.5 Mb inversion relative to B73.

The large inversion has now been reversed in one inbred



Creating inversions to preserve linkage blocks

Rönspies et al, 2022

If inversions can be reversed, they can also be created, thus forming permanent linkage blocks without recombination, but for recombination from DCOs in the loop (visible in the 10-14 Mb segment on the figure on the left below.



TRANSLOCATIONS or INTERCHANGES

Exchanges between non-homologous chromosomes

• Three types







Simple translocations are rare

 Normally need broken ends to bind with a fragment

Semisterility in Florida velvet bean Belling 1914/1915

Working with the Florida velvet bean, *Stizolobium deeringianum = Mucuna pruriens*

- Found plants with 50% aborted pollen and 50% seed set -- called this semisterility
- Half of offspring were normal, half were semisterile.



https://www.etsy.com/listing/635026498/muc una-pruriens-velvet-bean-cowitch-raw

Postulated 2 genes, K and L, such that

• Presence of K or L gave fertility





In 1920, moved to Cold Spring Harbor, and started working on Jimson weed plants ('B' white) that were not breeding in a Mendelian manner

Belling & Blakeslee 1924; 1925

- 1924: Belling & Blakeslee concluded that non-homologous chromosomes could exchange segments
- 1925: Associated translocations with cause of semisterility

Blakeslee 1928

Found a ring of 4 (\odot 4) in crosses of 'B' white x other plants of *Datura* (Jimson weed)

1926-1930: Translocations in Drosophila by Stern, Painter, and Muller



Semisterility in corn

Brink, 1927; Brink & Burnham, 1929

Find semisterility in corn (the same year McClintock karyotyped maize)





Burnham, 1930

Finds a \odot 4

McClintock, 1930

Identified cross-shaped configuration at pachytene



Meiosis in translocation heterozygotes graphics from Strickberger, 1976

Normal \times translocation homozygote to give a translocation heterozygote. Forms cross @ pachytene.



Case 1 – Crossovers in the Arms

Adjacent 1 separation

Non-sister centromeres go to the same pole & end up in separate daughter cells)

• Leads to formation of non-functional (duplicatedeficient) gametes









Adjacent 2 separation

Sister centromeres go to the same pole & end up in the same daughter cell

• Gametes formed are also duplicate-deficient







Alternate separation

Sister centromeres end up in the same daughter cell

- Gives a figure 8 configuration
- All gametes are functional
- Always recover #1 and #2 together
 - \circ There is no independent assortment





Translocation separations

If all three configurations occurred with equal frequency, $^2/_3$ of gametes would be nonviable. However, this is not observed.

Туре:	Expected	Drosophila	Maize	Barley
Adjacent 1	¹ / ₃	30	30	30
Adjacent 2	¹ / ₃	20	20	-
Alternate	¹ / ₃	50	50	70

Therefore, obviously, alternate separation takes place preferentially

Pseudolinkage

From Sturtevant & Beadle, 1939



Apparent linkage of genes on nonhomologous chromosomes

• Result of negation of independent assortment

All genes on the 2 chromosomes involved in a translocation behave as a single linkage group

The only phenotypes recovered are the double recessive (*aabb*) or the double dominants (*AB*).

Dominant/recessive combinations (*Abb* or *aaB*) are never recovered. I.e., the effects of independent assortment are negated



Effect on linkage maps

Homozygote: segments of the maps will be interchanged, reflecting change in the physical order of the genes on the chromosomes

Heterozygotes: gives a 4-armed map corresponding to the pachytene configuration

Effect of crossover number

A crossover event in each of the 4 arms of a cross gives a $\odot 4$



Two rings of 4 in barley, one undergoing adjacent (left) and the other alternate (center) separation. From **Haabera. 1960**.

A crossover in 3 of the 4 arms gives a chain of 4



Two chains of 4 in Agrostis. After Jones, 1956.

Case 2 – Crossovers in the interstitial region

I.e., the region between the centromere and the exchange point



Adjacent 1 separation



Net result is that 50% of gametes are functional.

• These result from the crossover event.

Adjacent 2 separation

Will not occur, or is rare, its occurrence depends on length of the interstitial region

Alternate



Net result is 50% of gametes are non-functional.

• These result from the crossover

Relative frequency of disjunctions worked out by McClintock (unpublished) and Burnham, 1950, using the NOR (chromosome 6) as a marker.

	Alternate	Adjacent 1	Adjacent 2
Short interstitial	50-57%	19-31%	19-26%
Long interstitial	55-56%	41-45%	0-3%

Effect of Translocations on recombination

Given the T5-9a translocation (i.e., the first translocation found between chromosomes 5 and 9) of maize:



Chromosome 5 (R) and part of chromosome 9 (L) showing break points. Horizontal arrows indicate regions showing variable pairing.

Chromosome	Standard	Heterozygous T5-9a as male	Homozygous T5-9a
Chromosome 9			
yg₂-sh	23	11	
sh-wx	20	5	18.6
WX-V ₁	12	11	independent
Chromosome 5			
bm-pr	27	32	
pr-wx	independent	28	23.8

The tendency is for a reduction in crossover frequency

In addition, whenever crossing over takes place in the interstitial region, alternate disjunction results in unviable gametes.

This leads to an overall decrease in the number of crossovers recovered, compressing the map distance

Frequency of crossing over in the interstitial region

 Worked out by McClintock (unpublished) and Burnham, 1950, using the NOR (chromosome 6) at the tetrad stage and pollen abortion as markers in translocations between chromosomes 6 & 5



Pachytene configuration of a 5-6 translocation. The interstitial regions are denoted as a and b. (Burnham, 1960).

- Number of NOR = number of nucleoli
- If no NOR is present, nucleolar material remains scattered in small droplets

IF NO CROSSING OVER OCCURS:

1) Adjacent 1:

- 2 have 2 nucleoli
- 2 have diffuse nucleolus
- All abort

2) Adjacent 2:

- All microspores have 1 nucleolus
- All abort



3) Alternate:

- All have 1 nucleolus
- All are viable

- Can't tell from adjacent 2 by looking at tetrad



Ŧ

2

4

3



IF A SINGLE CROSSOVER TAKES PLACE AT a:



Problem: Cannot tell adjacent 1 apart from alternate, as both tetrads are the same.

-The # of tetrads with 1 diffuse nucleolus gives the # of crossover tetrads

- The # of NCO separations is made of 3 types:

- Adjacent 1 = frequency of tetrads with 2 diffuse nucleoli

- Alternate = amount of viable pollen [alternate] - viable pollen from crossover tetrads [adj1 with

CO + alternate dis] = % tetrads with 1 diffuse × 2

- Adjacent 2 = amount of aborted pollen - adjacent 1 - aborted pollen from crossover

Permanent translocation heterozygotes

Gates 1908; Golczyk, Massouh & Greiner, 2014

(see section 5d on *Oenothera* cytogenetics)





Oenothera

It is possible to get every chromosome involved in a translocation

Originally found in *Oenothera* by Gates, 1908





Use of translocations in mapping & breeding

Burnham 1946, Sisodia & Shebeski, 1965; review by Farré et al., 2014

- Used for chromosome mapping, assigning linkage groups to chromosomes
- Used for development of physical maps
- Burnham (1946) tried to get *Oenothera*-type translocation complexes in maize and barley, but failed
 - Benefits: could get true-breeding hybrids from seed!
 - o As translocations increased, fertility decreased
 - Sisodia & Shebeski. 1965. Got ring chromosomes, but plants were sterile



- Works in *Oenothera*, but only because it is in conjunction with lethals, etc.
 - **Golczyk et al, 2014:** The end-segments are deprived of canonical telomeres but capped with constitutive heterochromatin
- Works better with metacentric chromosomes
- Also found in *Rhoeo discolor, Paeonia californica, Viscum fischeri,* and *V. engleri* (East African mistletoes).
 - In these mistletoes, 2n = 23 , 22 ♀.
 - The ♂ has 4 X and 5 Y chromosomes, all involved in translocations with the autosomes, so its gametes have 12 chromosomes.
 - \circ The ^Q has 8 X chromosomes, none of which are involved in translocations.
 - It forms 11 II, and its gametes have 11 chromosomes.