

Inversions

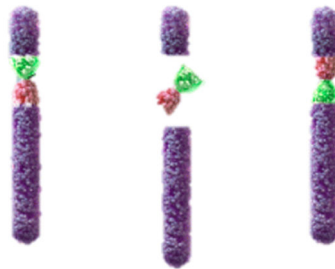
Muller, 1916, working with *Drosophila*, found C (for crossover reducers) factors reduced or suppressed

A paracentric inversion



From US National Library of Medicine

A pericentric inversion



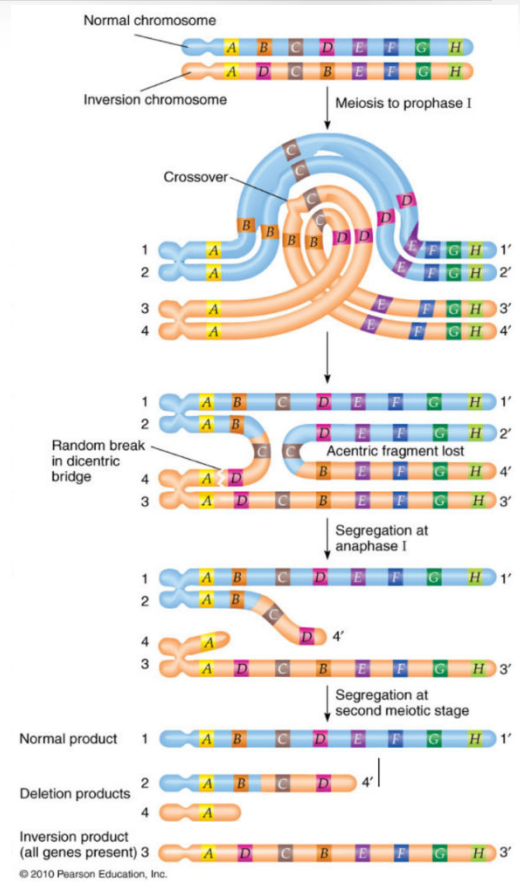
Paracentric inversions



Russell & Burnham 1950

Crossover suppression

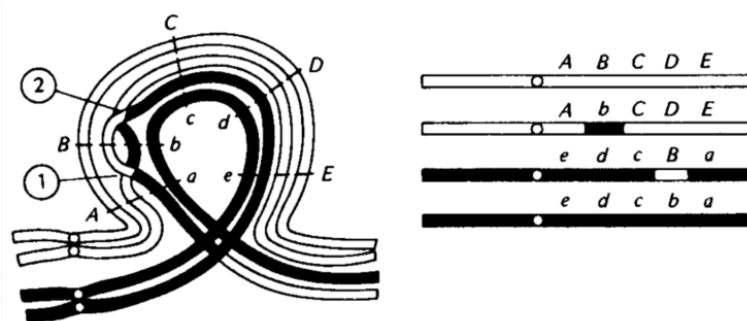
McClintock, 1931; 1933



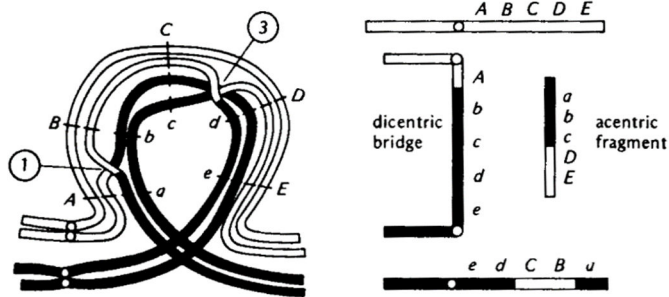
Double Crossovers in the Inversion Loops

graphics from Strickberger, 1976

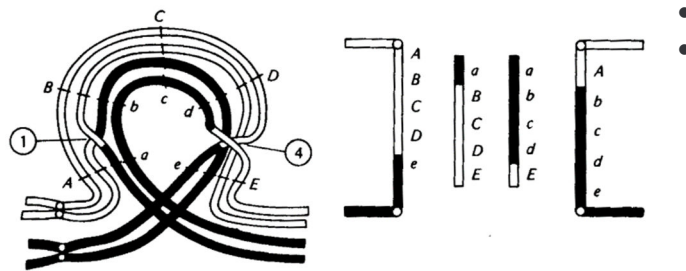
1) 2-strand double crossover



2) 3-strand double crossover



3) 4-strand double crossover



If one of the double crossovers occurs outside of the loop

Modified from **McClintock, 1938**

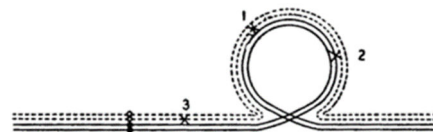
A. Get same configuration for a 1 or 3 strand double crossover

- Not affected by CO outside of inversion area

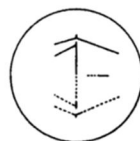
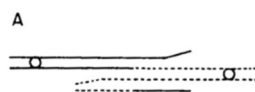
From **Burnham, 1962**, after **McClintock, 1938**

Ana I from Stevens & Bougourd. 1991. Heredity 66: 391-401

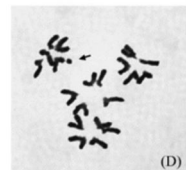
Ana II from Wang & Zhang. 2007. Plant Sci 172: 380-392



ANA I



ANA II

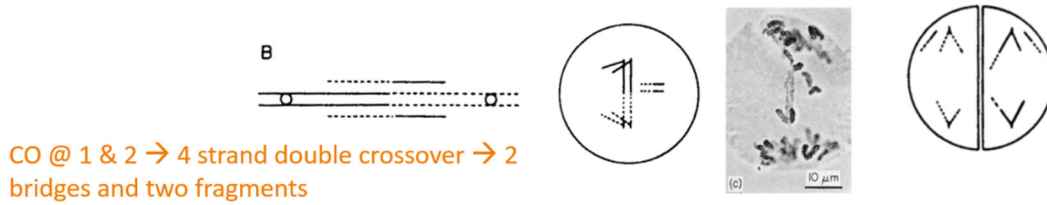


Single Crossover @ 1 or 2 → bridge & fragment at anaphase I



(D)

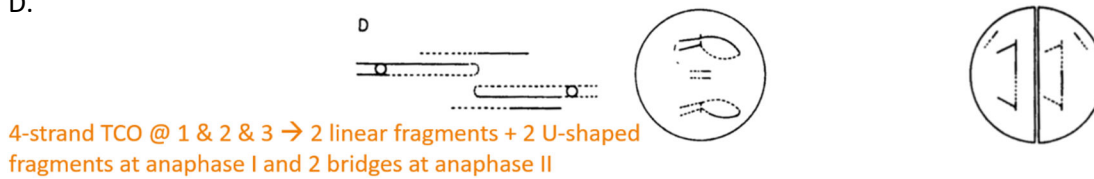
B. Configuration of 4-strand double crossover



C.



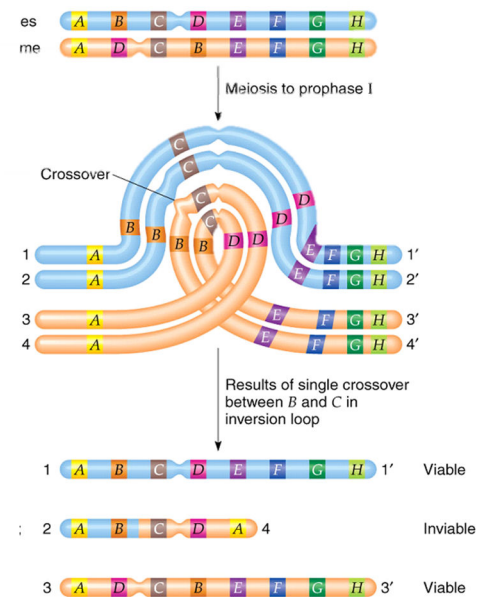
D.



Pericentric inversions

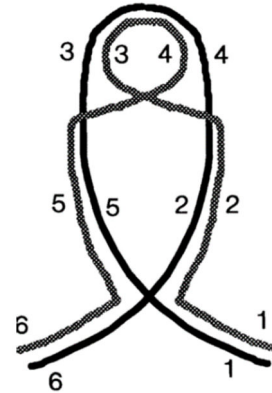
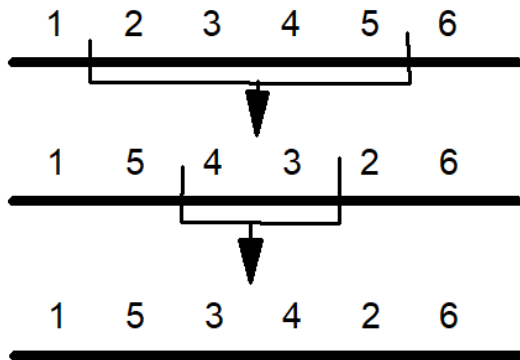


McClintock, 1931

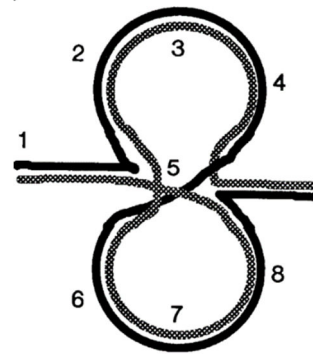
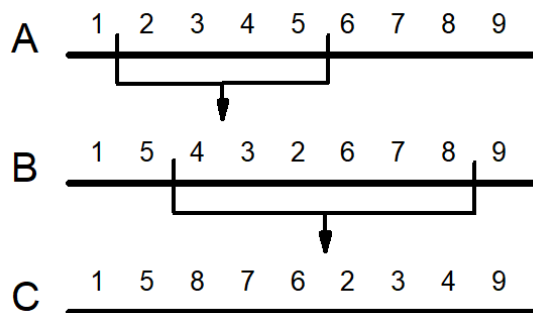


Multiple inversions

Case 1: Nested inversions



Case 2: Overlapping inversions

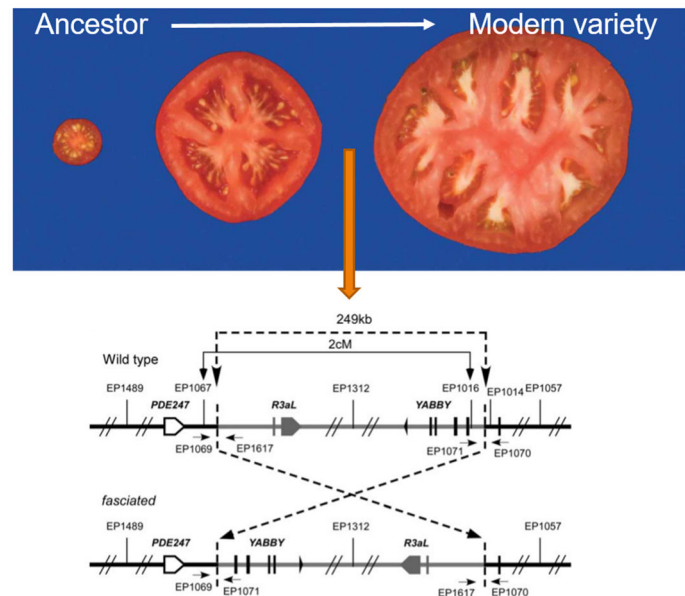


Role of inversions

Dobzhanski, with *Drosophila*:

Structural variants - inversions

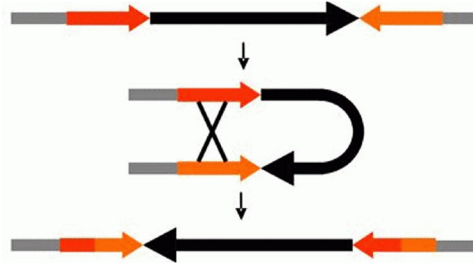
Huang & van der Knaap, 2011



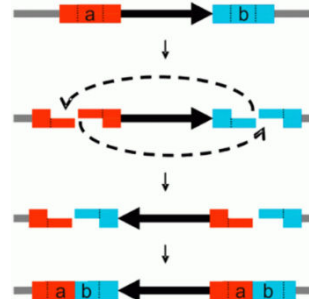
Origin of inversions

Casals & Navarro, 2007

Model 1: Duplications lead to inversions



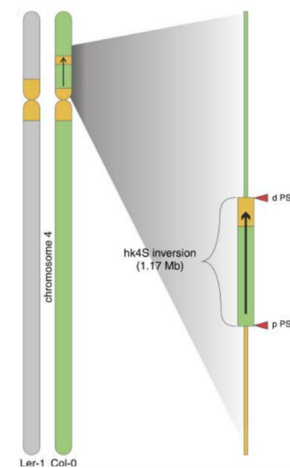
Model 2: Staggered breaks



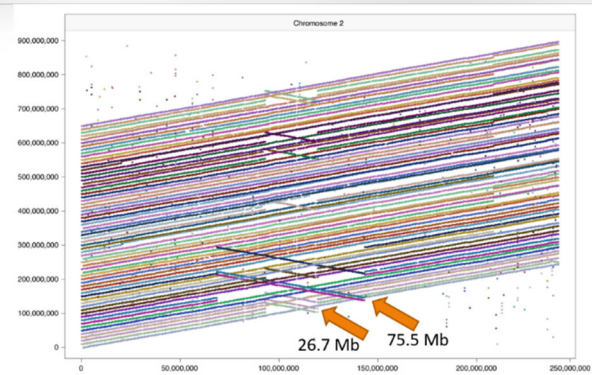
Reversing inversions

Schmidt et al, 2020

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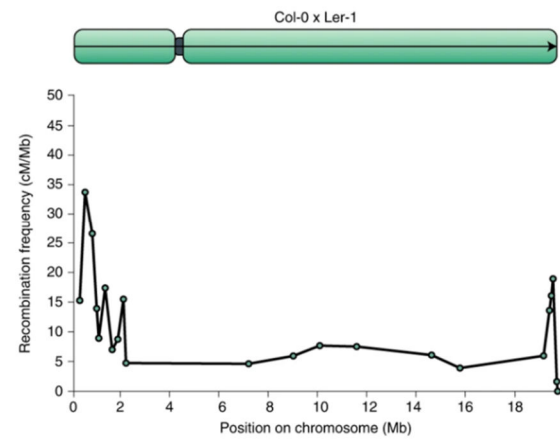
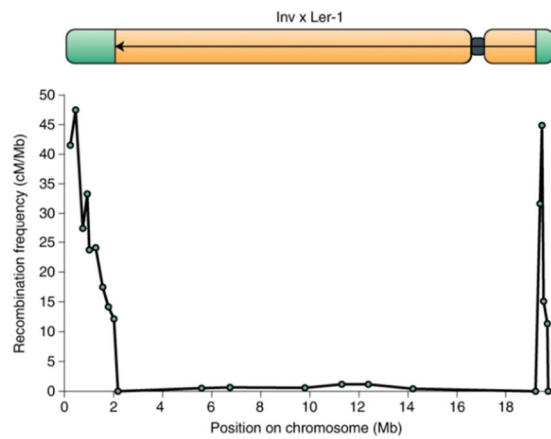


Schwartz et al, 2020



Creating inversions to preserve linkage blocks

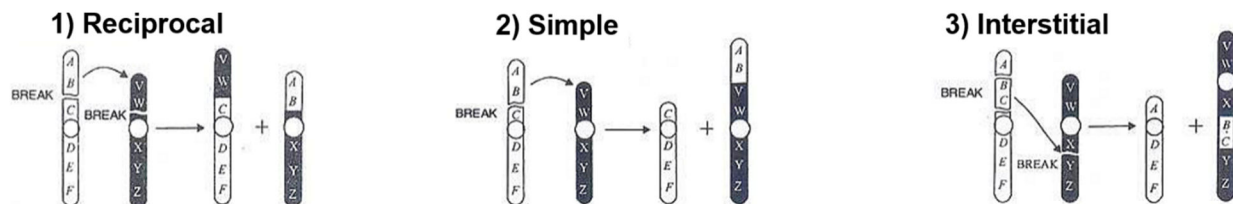
Rönspies et al, 2022



TRANSLOCATIONS or INTERCHANGES

Exchanges between non-homologous chromosomes

- Three types

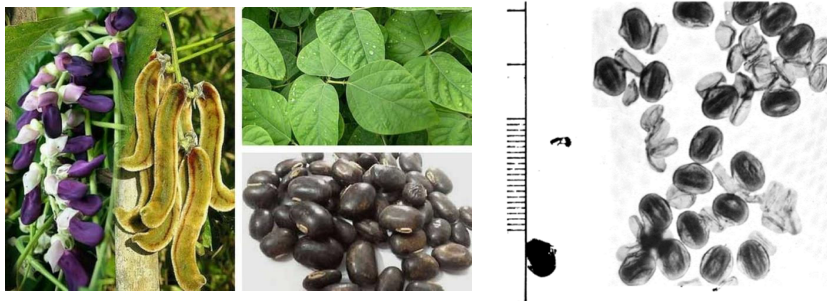


- Simple translocations are rare
 - Normally need broken ends to bind with a fragment

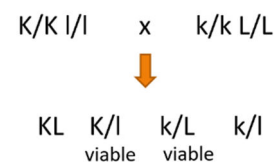
Semisterility in Florida velvet bean

Belling 1914/1915

Working with the Florida velvet bean, *Stizolobium deeringianum* = *Mucuna pruriens*



<https://www.etsy.com/listing/635026498/mucuna-pruriens-velvet-bean-cowitch-raw>



Postulated 2 genes, K and L, such that

- Presence of K or L gave fertility

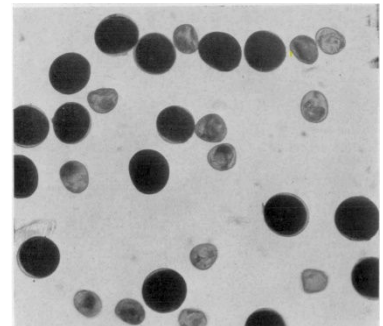
Belling & Blakeslee 1924; 1925



Blakeslee 1928

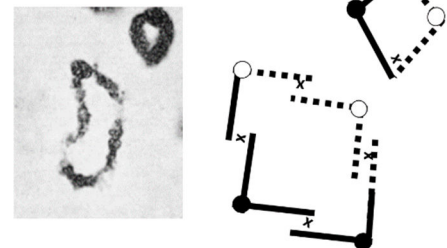
Semisterility in corn

Brink, 1927; Brink & Burnham, 1929

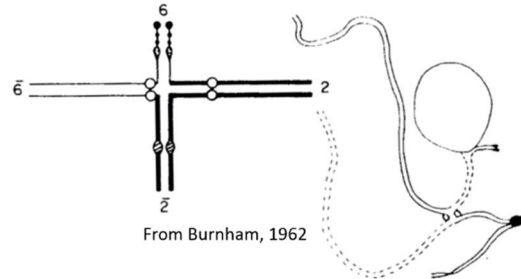
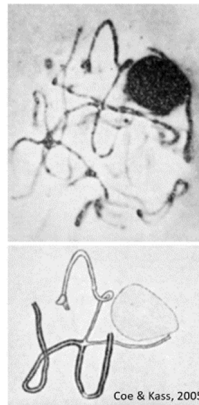


Burnham, 1930

A ring of 4 in maize.
Coe & Cass, 2005



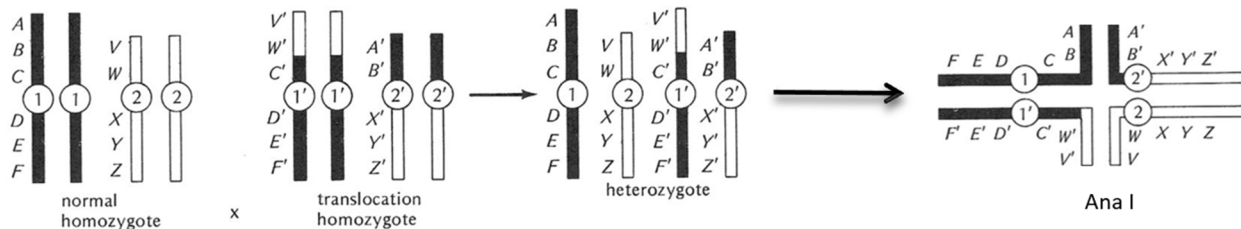
McClintock, 1930



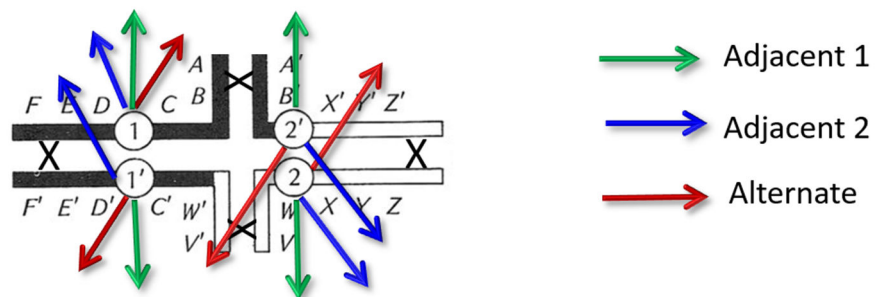
Meiosis in translocation heterozygotes

graphics from Strickberger, 1976

Normal × translocation homozygote to give a translocation heterozygote. Forms cross @ pachytene.



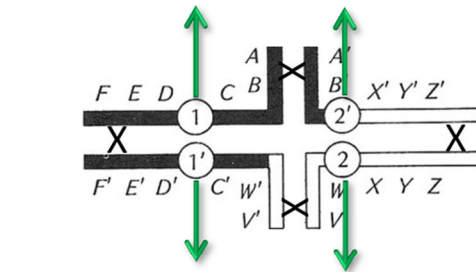
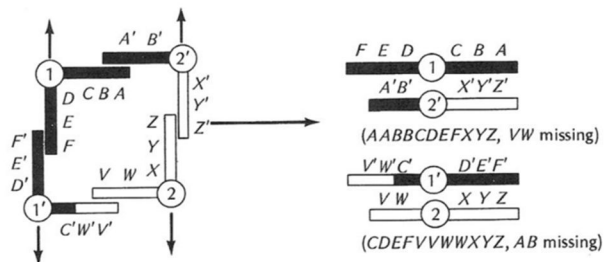
3 possible disjunctions



Case 1 – Crossovers in the Arms

Adjacent 1 separation

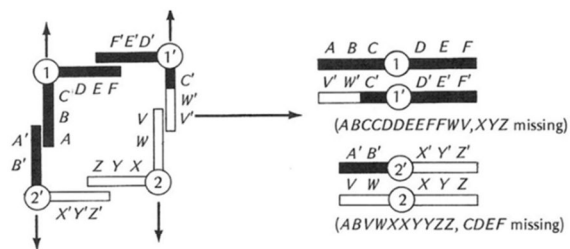
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-
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➔ All gametes abort

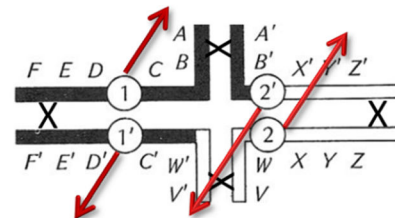
Adjacent 2 separation

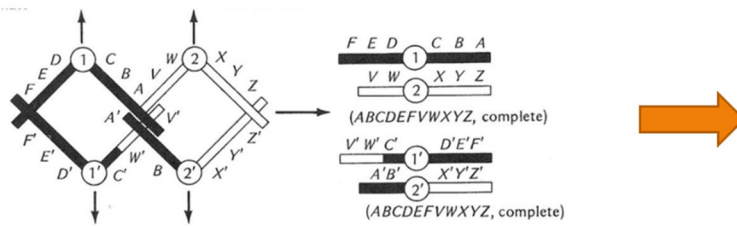
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➔ All gametes abort

Alternate separation



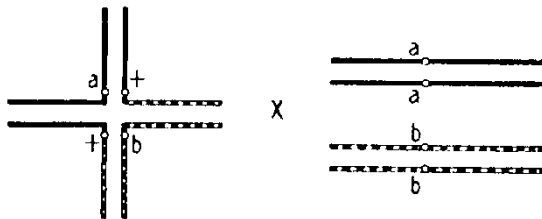


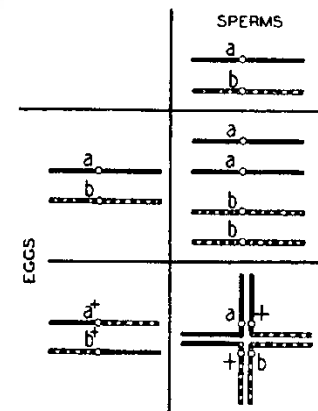
Translocation separations

Type:	Expected	Drosophila	Maize	Barley
Adjacent 1	$\frac{1}{3}$	30	30	30
Adjacent 2	$\frac{1}{3}$	20	20	-
Alternate	$\frac{1}{3}$	50	50	70

Pseudolinkage

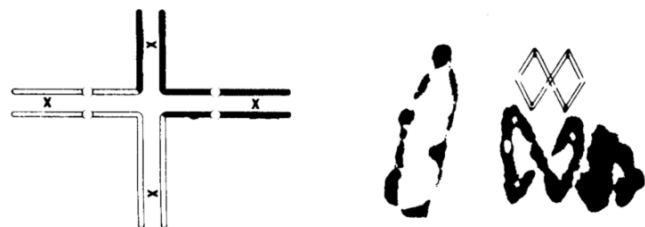
From Sturtevant & Beadle, 1939



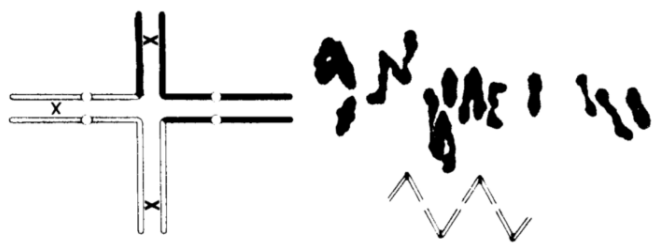


Effect on linkage maps

Effect of crossover number

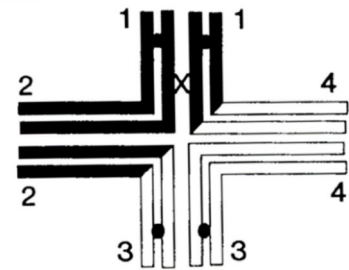


Two rings of 4 in barley, one undergoing adjacent (left) and the other alternate (center) separation. From Hagberg, 1960.

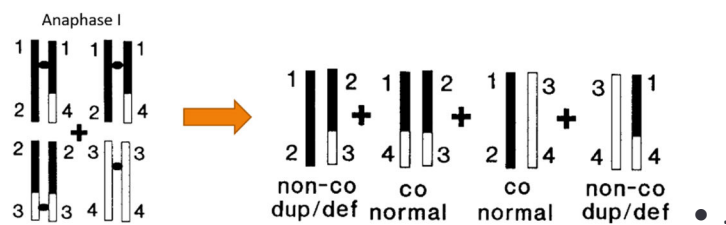


Two chains of 4 in Agrostis. After Jones, 1956.

Case 2 – Crossovers in the interstitial region



Adjacent 1 separation



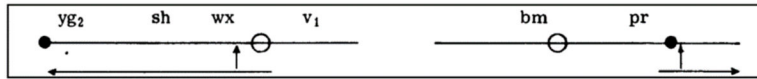
Adjacent 2 separation

Alternate



	Alternate	Adjacent 1	Adjacent 2
Short interstitial	50-57%	19-31%	19-26%
Long interstitial	55-56%	41-45%	0-3%

Effect of Translocations on recombination

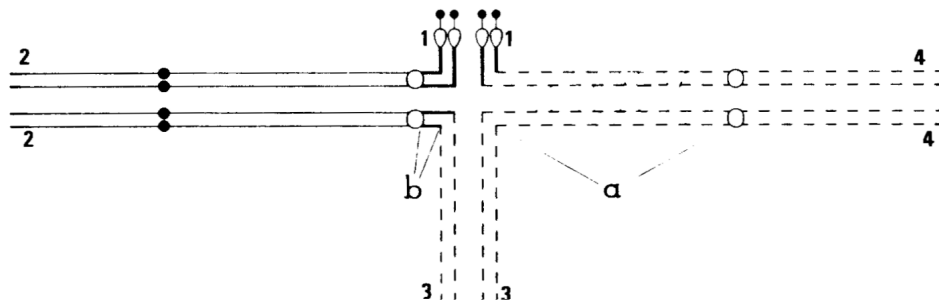


Chromosome 5 (R) and part of chromosome 9 (L) showing break points.
Horizontal arrows indicate regions showing variable pairing.

Chromosome	Standard	Heterozygous T5-9a as male	Homozygous T5-9a
Chromosome 9			
yg_2-sh	23	11	--
$sh-wx$	20	5	18.6
$wx-v_1$	12	11	independent
Chromosome 5			
$bm-pr$	27	32	--
$pr-wx$	independent	28	23.8

Frequency of crossing over in the interstitial region

- Worked out by McClintock (unpublished) and Burnham, 1950, using the NOR (chromosome 6) at the tetrad stage and pollen abortion as markers in translocations between chromosomes 6 & 5



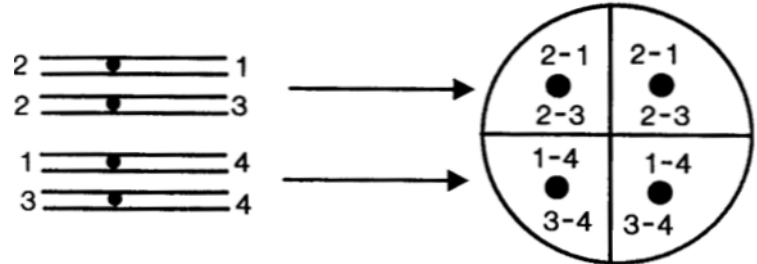
Pachytene configuration of a 5-6 translocation. The interstitial regions are denoted as a and b. (Burnham, 1960).

- Number of NOR = number of nucleoli
- If no NOR is present, nucleolar material remains scattered in small droplets

IF NO CROSSING OVER OCCURS:

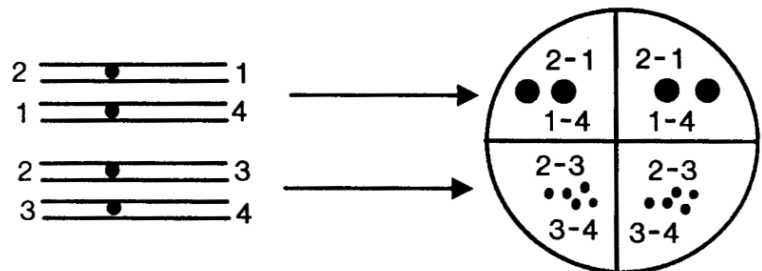
1) Adjacent 1:

- 2 have 2 nucleoli
- 2 have diffuse nucleolus
- All abort



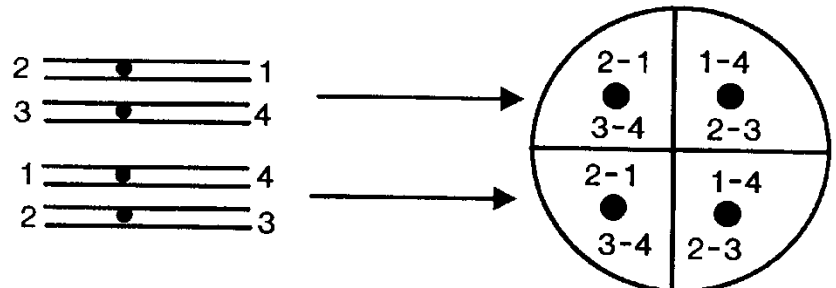
2) Adjacent 2:

- All microspores have 1 nucleolus
- All abort



3) Alternate:

- All have 1 nucleolus
- All are viable
- Can't tell from adjacent 2 by looking at tetrad



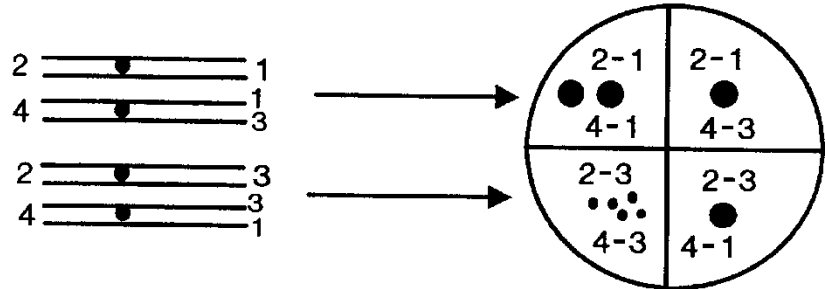
IF A SINGLE CROSSOVER TAKES PLACE AT a:

Adjacent 2 - will not occur

Adjacent 1:

- 2 are viable

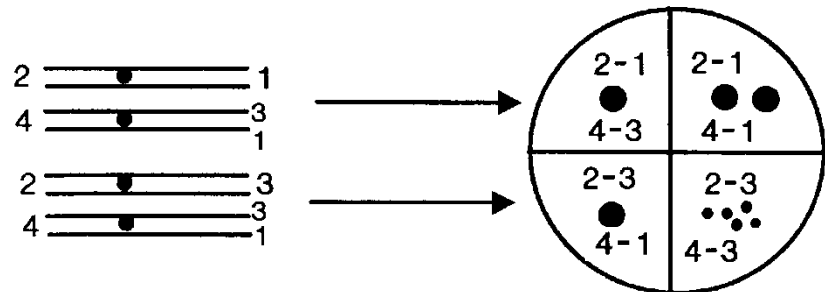
- 2 abort (1 diffuse)



Alternate:

- 2 viable

- 2 abort (1 diffuse)



Problem: Cannot tell adjacent 1 apart from alternate, as both tetrads are the same.

-The # of tetrads with 1 diffuse nucleolus gives the # of crossover tetrads

- The # of NCO separations is made of 3 types:

- Adjacent 1 = frequency of tetrads with 2 diffuse nucleoli

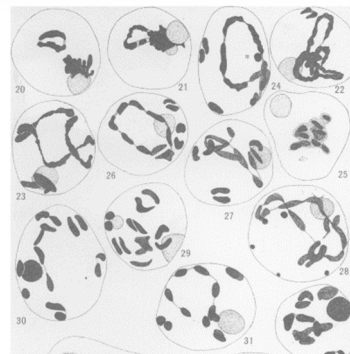
- Alternate = amount of viable pollen [alternate] - viable pollen from crossover tetrads [adj1 with CO + alternate dis] = % tetrads with 1 diffuse × 2

- Adjacent 2 = amount of aborted pollen - adjacent 1 - aborted pollen from crossover

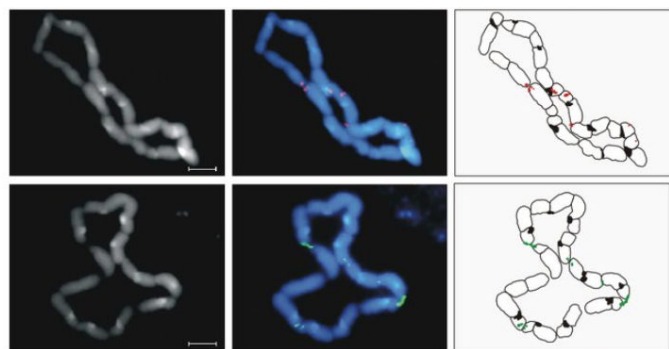
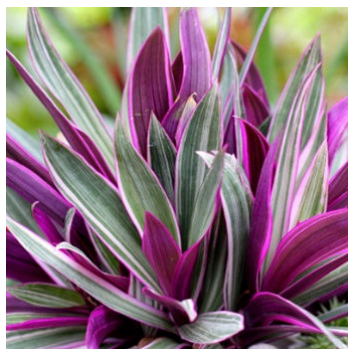
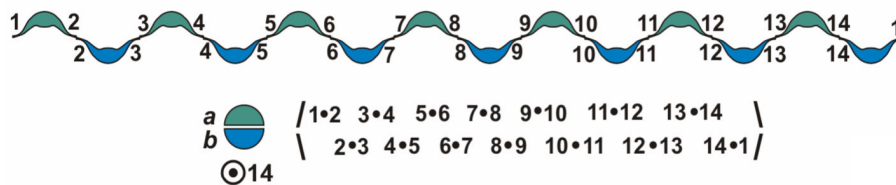
Permanent translocation heterozygotes

Gates 1908; Golczyk, Massouh & Greiner, 2014

(see section 5d on *Oenothera* cytogenetics)



Oenothera



Use of translocations in mapping & breeding

Burnham 1946, Sisodia & Shebeski, 1965; review by Farré et al., 2014



© 14 in barley, Sisodia & Shebeski, 1965

- Also found in *Rhoeo discolor*, *Paeonia californica*, *Viscum fischeri*, and *V. engleri* (East African mistletoes).
 - In these mistletoes, $2n = 23\sigma, 22\varphi$.
 - The σ has 4 X and 5 Y chromosomes, all involved in translocations with the autosomes, so its gametes have 12 chromosomes.
 - The φ has 8 X chromosomes, none of which are involved in translocations.
 - It forms 11 II, and its gametes have 11 chromosomes.